

Introduction

Long mate-pair libraries are invaluable tools for genome assembly. However, traditional methods of long mate-pair library construction require large (20 µg) quantities of DNA and several days of hands-on time. Illumina's Nextera™ Long Mate-Pair (LMP) method is rapid and requires only 1 to 4 micrograms of input material. Here we present an initial assessment of the method for both gel-free and gel size-selected libraries using microbial, fungal, and plant samples. We observed uniform read coverage and high read uniqueness for Nextera™ LMP libraries. Assembly using ALLPATHS-LG generated low contig and scaffold numbers even with relatively low mate-pair coverage.

Methods Overview

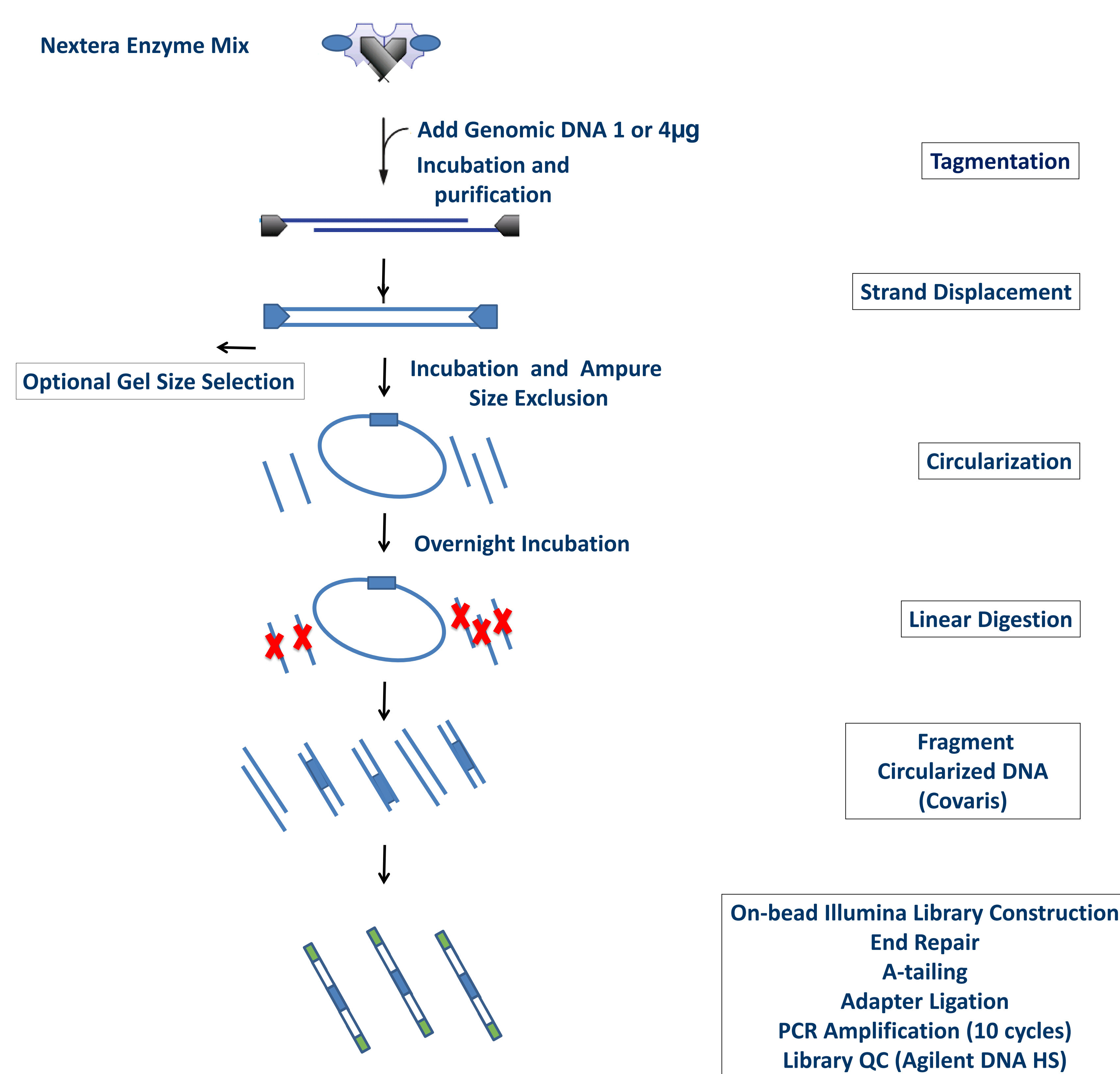
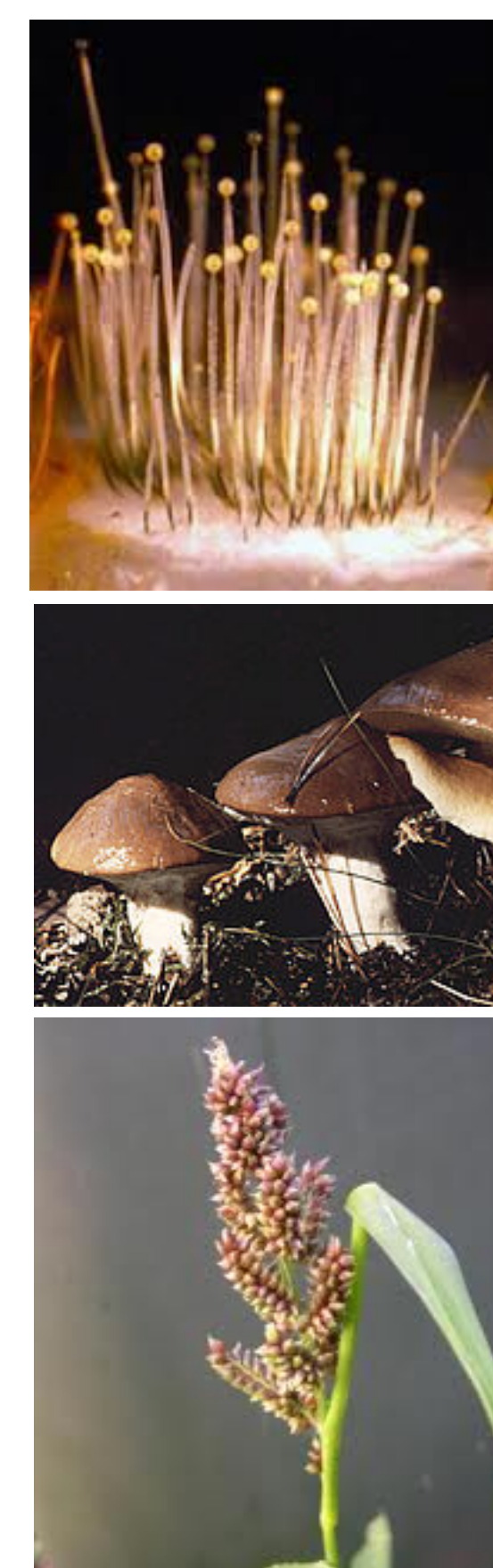


Figure 1. Nextera LMP Workflow

Organisms Tested

Species	%GC	Type
<i>Phycomyces blakesleeanus</i>	36%	Filamentous fungi
<i>Spirochaeta smaragdinae</i>	49%	Gram (-) microbe
<i>Conexibacter woesei</i>	73%	Gram (+) microbe
<i>Cellomonas flavigena</i>	74%	Gram (+) microbe
<i>Suillus luteus</i>	47%	Basidiomycete fungi
<i>Sorghum bicolor</i>	42%	Plant

Table 1. Initial testing organisms and their GC-content



Results

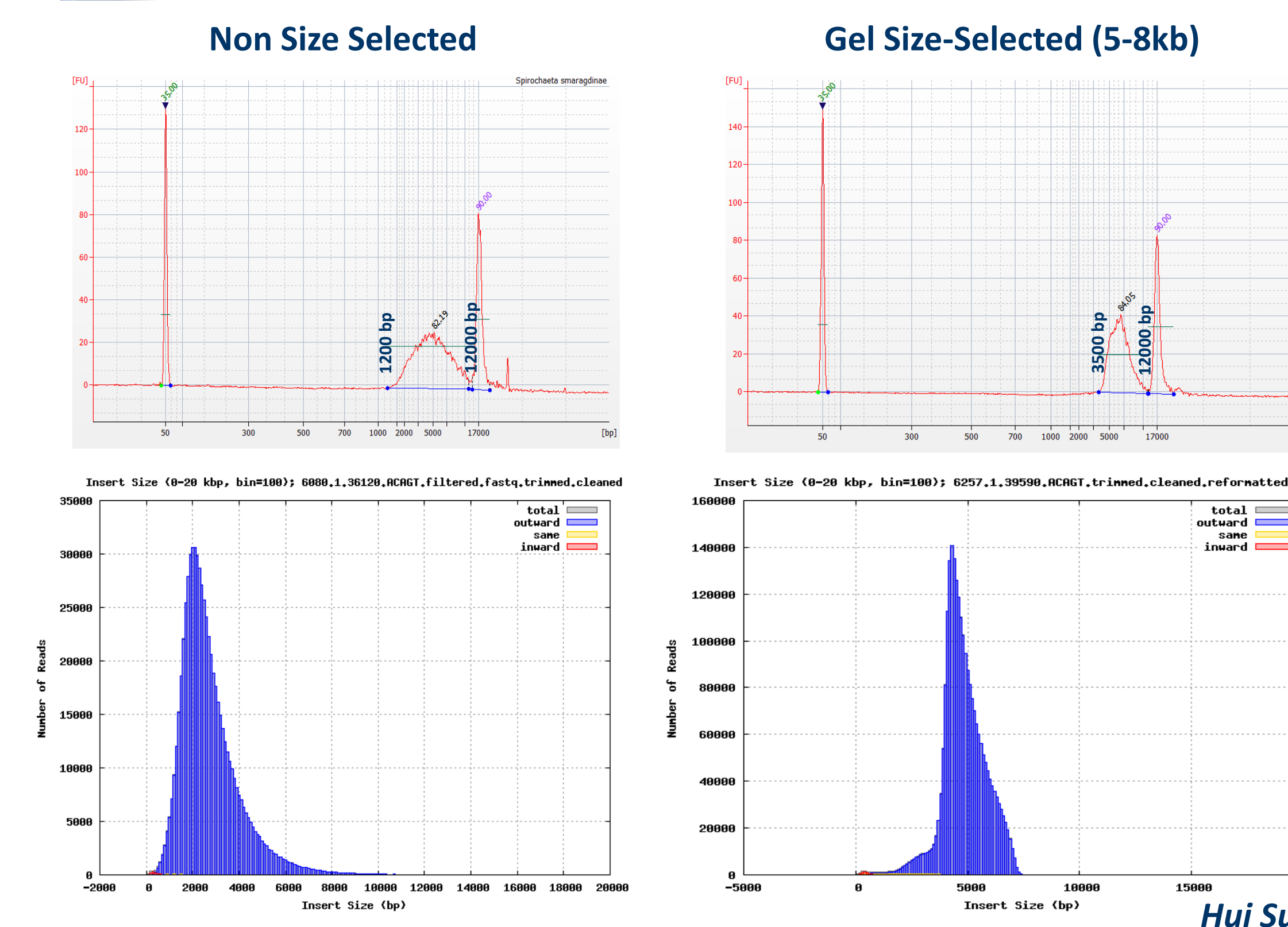


Figure 2. Size distribution of gel-free and gel size-selected libraries

Species	% Mapped Reads Non size-selected	% Mapped Reads Gel size-selected
<i>Phycomyces blakesleeanus</i>	96%	96%
<i>Spirochaeta smaragdinae</i>	97%	98%
<i>Conexibacter woesei</i>	88%	NA
<i>Cellomonas flavigena</i>	88%	94%
<i>Suillus luteus</i>	71%	NA
<i>Sorghum bicolor</i>	94%	NA

Table 2. Nextera LMP yields high percentage of mapped reads.

Organism & Assembly Type	Scaffolds	Contigs	Scaffold L50	Contig L50
<i>Conexibacter woesei</i> Frag+Traditional LMP	1	7	6355 Kb	1190 Kb
<i>Conexibacter woesei</i> Frag+ Nextera LMP	1	8	6328 Kb	744 Kb
<i>Cellomonas flavigena</i> Frag+Traditional LMP	8	48	4060 Kb	188 Kb
<i>Cellomonas flavigena</i> Frag+ Nextera LMP	4	27	3493 Kb	408 Kb
<i>Suillus luteus</i> Fragment only	1944	2113	57.6 Kb	51.3 Kb
<i>Suillus luteus</i> Frag+ Nextera LMP	397	1477	240 Kb	54.6 Kb

Table 3. ALLPATHS-LG assemblies were improved with the inclusion of Nextera LMP data compared to traditional LMP data.

Summary

- User-friendly protocol with short hands-on time
- Low template requirement compared to traditional long-mate pair methods (1µg/4 µg)
- Read uniqueness is high for Nextera LMP libraries
- Nextera LMP libraries have uniform read coverage
- Insert size doesn't seem to have significant impact on contig N50
- ALLPATHS-LG generated low contig and scaffold numbers for microbes, even with low coverage
- Addition of Nextera LMP data generally improved assembly results

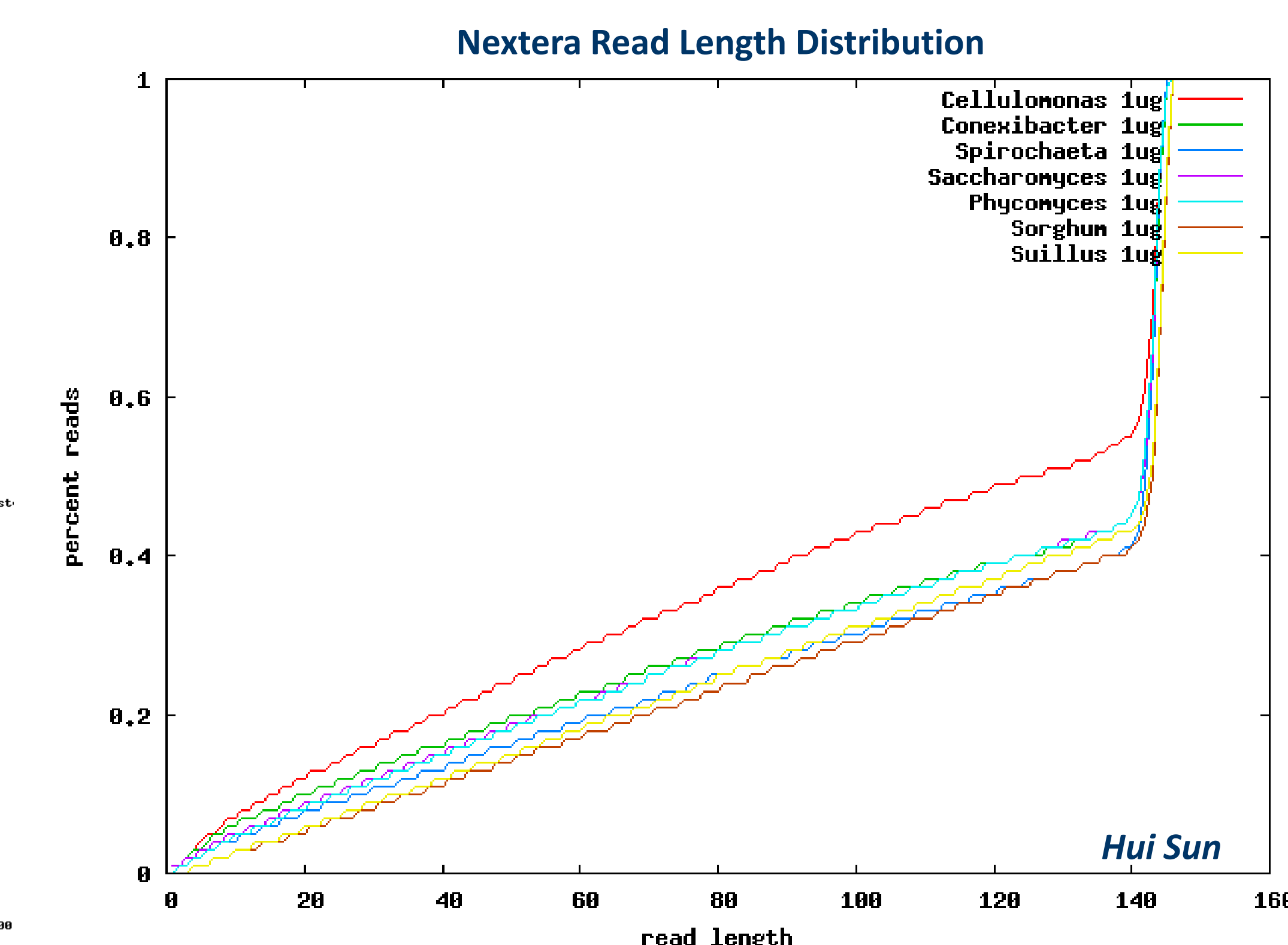


Figure 3. Uniform line indicates that Nextera transposon is inserted randomly across entire read length distribution.

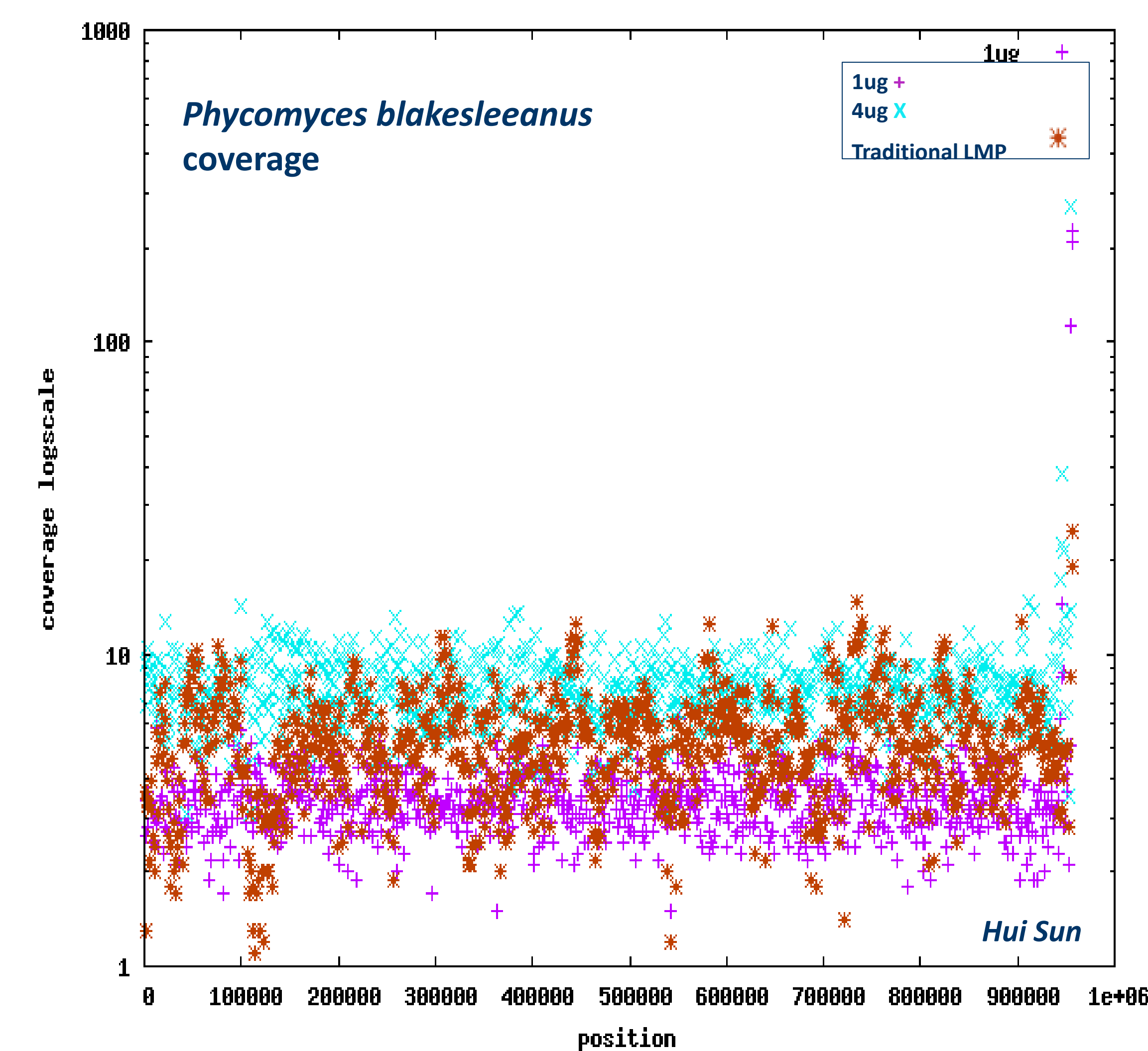


Figure 4. Nextera LMP coverage across scaffold is more evenly distributed compared to the traditional LMP library.